A poorly differentiated pulmonary squamous cell carcinoma in a free-ranging
Atlantic bottlenose dolphin (Tursiops truncatus)

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Abstract. A free-ranging, adult, female offshore bottlenose dolphin (Tursiops truncatus) was found freshly
dead in 1999 on Ocean Park Beach in San Juan, Puerto Rico. The left-lung and right-lung pleura had multiple
white, firm-to-hard nodules with coagulative necrosis. Histologically, the neoplasms were characterized by
multiple well-circumscribed, nonencapsulated expansile masses consisting mostly of polygonal cells with fewer
circumferential flattened basoid cells that compressed alveoli, bronchioles, and bronchi. Neoplastic cells
stained positive for cytokeratin, with sporadic vimentin staining, and were negative for epithelial membrane
antigen, thyroid transcription factor-1, calretinin, and human mesothelial cell antigen. A diagnosis of poorly
differentiated pulmonary squamous cell carcinoma with lymph node and renal metastases was made on the
basis of histomorphology and immunohistochemical staining. This is the first documentation of pulmonary
squamous cell carcinoma in a dolphin.

Reported neoplasms of the respiratory tract in odontocetes include reticuloendotheliosis in a bottlenose
dolphin and squamous cell carcinoma in an Amazon porpoise.15 Few neoplasms have been reported in bot-
tlenose dolphins, including renal adenoma, lymphoma, squamous cell carcinoma, and pancreatic carcino-
ma.15,15,16,18 A severely emaciated, free-ranging, adult, female offshore Atlantic bottlenose dolphin (Tursiops
truncatus) was found freshly dead on July 4, 1999, on Ocean Park Beach in San Juan, Puerto Rico. On the
basis of histopathology and immunohistochemistry, a diagnosis of poorly differentiated pulmonary carcino-
ma with lymph node and renal metastases was made. This report describes the morphologic and immuno-
histochemical features of the first reported poorly differentiated pulmonary squamous cell carcinoma with
renal and lymph node metastases in a stranded bottlenose dolphin.

The dolphin was found dead on Ocean Park Beach in San Juan, Puerto Rico (18°27.9′N, 66°03.1′W). Mitochon-
drial DNA sequence analysis performed on skin samples indicated that this dolphin was of the offshore
(vs. coastal) Tursiops morphotype (P. Rosel, unpublished data). Members of the Caribbean Stranding Net-
work of the Southeastern Marine Mammal Stranding Network conducted the necropsy in accordance with
systems were dissected, sampled, and preserved in 10% neutral buffered formalin. Specimens were pro-
cessed routinely for histopathology into paraffin blocks, sectioned at 4–5 μm, and stained with hema-
toxylin and eosin (HE). Sections of tumor were stained for vimentin (VIM, 1:1,500), human mesothelial cell
antigen (HBME-1, 1:50), keratin AE1/AE3 (1:200), high molecular weight keratin (1:50), epithelial mem-
brane antigen (EMA, 1:1,500), thyroid transcription factor-1a (TTF-1, 1:150), keratin CAM5.2b (1:1,500),
and calretinin (CALRET, 1:200) by immunohistochemistry. The cytokeratin antibodies were combined
at the aforementioned dilutions for use, a standard technique followed in the University of Miami, School of
Medicine, Pathology Reference Service, Immunoperoxidase Laboratory, Miami, Florida. A mixture of
sera from nonimmune normal mouse, rabbit, and goat was applied in lieu of the primary antiserum for nega-
tive antibody controls. Normal thyroid and lung sections from another offshore bottlenose dolphin were
stained as positive controls for TTF-1. Normal splenic

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serosal mesothelium from a bottlenose dolphin was stained as control for CALRET and HBME-1. The CALRET was immunoreactive, whereas the HBME-1 was not. Routine human known positive controls were used for all other immunohistochemical stains (M. Nadji, personal communication).

The dolphin was 273 cm in length from rostrum to tail notch and weighed 149.1 kg. Grossly, the dolphin was severely emaciated with protruding scapulae, ribs, and vertebral transverse processes. The lungs and major bronchi contained frothy white fluid and were diffusely congested. Most of the left lung was diffusely and severely affected with multiple irregular, round-to-oval, white, firm-to-hard areas of approximately 2–3 cm with coagulative necrosis. There were similar smaller discrete nodules along the visceral pleura of the right lung. All thoracic lymph nodes were enlarged and mottled white and maroon on cut section.

Histologically, the pulmonary parenchyma contained multiple well-circumscribed, nonencapsulated expansile masses, which compressed and/or infiltrated alveoli, bronchioles, and bronchi. The neoplastic cells had a squamous morphology consisting of mostly polygonal cells, with a mantle of flattened basaloid cells (Fig. 1A). The polygonal cells had moderate eosinophilic cytoplasm, distinct cell borders, round-to-elongate hyperchromatic or vesicular nuclei, and occasional single prominent nucleoli (Fig. 1B). There was moderate variability in the sizes of cells and nuclei; mitoses were infrequent at 1–2 per high-power field. The cells were arranged in variably sized nests with frequent central necrosis, neutrophilic inflammatory cell infiltration, and a mild, thin, dense, fibrous stroma. The pleural lymphatic channels and blood vessels contained individual or clustered neoplastic cells.

The normal offshore bottlenose dolphin thyroid and lung sections had positive follicular and respiratory epithelial cell nuclear staining for TTF-1. Normal dolphin splenic serosal mesothelium stained positive for CALRET but was negative for HBME-1. The cytokeratin markers were diffusely and strongly positive in all neoplastic cells with peripheral cytoplasmic staining (Fig. 1C). Neoplastic cells were nonimmunoreactive for EMA, TTF-1, HBME-1, and CALRET and were sporadically positive for VIM.

The normal architecture of a thoracic lymph node was mostly effaced by similar neoplastic cells admixed with moderate, multifocal, neutrophilic inflammatory cell infiltration and necrotic cellular debris. A rimulus had a glomerular metastatic focus, which replaced the glomerular tuft and distended the Bowman's capsule. In addition, the renal metastatic focus was similarly immunoreactive for all cytokeratin markers used. Occasional keratin protein-reactive neoplastic cells were present within renal interlobular artery lumina.

Figure 1. A. Lung; bottlenose dolphin. Poorly differentiated carcinoma. Variously sized nests of polyhedral cells distorting bronchioles (Br) and alveoli (A) with frequent central necrosis and a thin, dense, fibrous stroma (arrows). HE. Bar = 200 μm. B. Lung; bottlenose dolphin. Poorly differentiated carcinoma. Neoplastic cells with moderate eosinophilic cytoplasm, round-to-elongate hyperchromatic or vesicular nuclei, distinct cell borders, anisokaryosis, anisocytosis, and occasional binucleation. HE. Bar = 10 μm. C. Lung; bottlenose dolphin. Poorly differentiated carcinoma. Neoplastic cells were diffusely and strongly immunoreactive for the combined cytokeratin antibodies with occasional marked peripheral staining. Avidin-biotin-diaminobenzidine-peroxidase complex method with hematoxylin counterstain. Bar = 50 μm.
The renal tubular epithelial cells had weak diffuse EMA cytoplasmic staining, whereas the metastatic focus was negative for EMA.

Despite the lack of classical characteristic features of a squamous cell carcinoma, the neoplasm morphology, degree of cellular anaplasia, and immunostaining characteristics were consistent with a diagnosis of poorly differentiated squamous cell carcinoma. Differential diagnoses included a primary bronchogenic carcinoma and mesothelioma. A metastatic epithelial cell tumor was considered to be unlikely because a larger primary neoplasm was not observed elsewhere in the carcass; however, an occult neoplasm could not be ruled out.

Epithelial membrane antigen and cytokeratin proteins are markers for epithelial differentiation; however, EMA is neither as sensitive nor as specific as keratin. Keratin proteins have strong immunoreactivity for squamous cell carcinoma and mesothelioma. In human surgical pathology, EMA is considered a marker for epithelial differentiation and is immunoreactive for adenocarcinomas of various primary locations; however, certain mesenchymal and lymphoreticular neoplasms also show immunoreactivity. There are also discrepancies regarding EMA reactivity in mesothelioma and squamous cell carcinoma between investigators reporting both positive and negative results.

Thyroid transcription factor-1 is a nuclear transcription protein and a tissue-specific factor that is expressed in human thyroid follicular cells, bronchial and alveolar epithelium, and the diencephalon. Immunoreactivity to TTF-1 has been used to distinguish positive primary pulmonary adenocarcinoma from metastatic adenocarcinoma and mesothelioma, which tend to be nonimmunoreactive (M. Nadji, personal communication). In humans, pulmonary squamous cell carcinomas are nonimmunoreactive for TTF-1 (M. Nadji, personal communication). The normal dolphin thyroid and lung sections had positive epithelial nuclear staining for TTF-1, supporting its use as a tissue-specific marker in dolphins. Nonspecific staining of mesenchymal cells, by TTF-1, in sections from the control and case specimens was not observed nor was there false-positive immunoreactivity for renal epithelial cells in the case specimen. The neoplastic cells in the lung, lymph node, and renal cortex were nonimunoreactive for TTF-1 and EMA but had strong keratin protein immunodetection.

Vimentin, HBME-1, and CALRET antibody stains were applied to differentiate carcinoma from malignant mesothelioma. Neoplastic cells were nonimmunoreactive for HBME-1 and CALRET and showed sporadic positive staining for VIM. Although there is controversy regarding definitive staining features of human mesotheliomas, most studies indicate that CALRET and HBME-1 are highly specific and sensitive markers for mesotheliomas. Normal mesothelial cells from a dolphin spleen stained positive for CALRET, confirming the use of this antibody in this species. However, HBME-1 antibody failed to stain normal dolphin splenic mesothelial cells; therefore, interpretation of this stain is limited. Vimentin has also been shown to positively stain human mesothelioma tumor cells; however, the frequency of positive staining is variable. In addition, VIM, typically a mesenchymal cell marker, has been shown to stain other carcinomas and may reflect epithelial–mesenchymal transition events. These findings lend support to ruling out malignant mesothelioma as a differential diagnosis for this neoplasm.

Primary pulmonary squamous cell carcinoma is relatively uncommon in nonhuman mammalian species. In humans, there is a positive correlation between lung cancer and tobacco smoking, chronic industrial hazard exposure, air pollution, molecular genetic alterations, and scar tissue. In humans, scar tissue is an antecedent to bronchogenic carcinoma and squamous cell carcinoma development and is also a desmoplastic response to the neoplasm. Exuberant or aberrant forms of inflammatory, reparative, and regenerative processes may serve as predisposing factors for oncogenesis.

Bronchial and bronchiolar mucosal mineralization, pulmonary granulomas, and verminous bronchopneumonia are not uncommon in both free-ranging offshore and coastal bottlenose dolphins (Ewing et al., unpublished data). Various genera of lung nematodes have been described in both coastal and pelagic small odontocetes. There is reported circumstantial evidence of prenatal Halocercus lagenorhynchus infection with transplacental transmission in the Atlantic bottlenose dolphin. Halocercus sp. have prominent cuticular spines that stimulate mucus production, marked acute inflammation, epithelial desquamation, hemorrhage, and Type II pneumocyte hyperplasia. These changes progress to pyogranulomas, histiocytic granulomas, reparative fibrosis, fibrous encapsulation, and mineralization that characterize the chronic phase of inflammation. Bronchiolar squamous metaplasia has not commonly been described as a characteristic of dolphin pulmonary nematodiasis; however, this was observed in an unrelated case of another offshore bottlenose dolphin with pulmonary halocerciasis (Ewing, unpublished data). In this species, persistent pulmonary nematodiasis may induce neoplastic transformation in metaplastic epithelium. Parasite-induced oncogenesis has been observed historically in rats, dogs, and humans associated with Cysticercus fuscricoloris, Spirocerca lupi, and Schistosoma haematobium, respectively.

Human papillomavirus–induced juvenile laryngotracheal and pulmonary papillomatosis with malignant
transformation can cause pulmonary squamous cell carcinoma. Papillomavirus-induced malignant transformation is recognized in cattle and is suspected in goats and rabbits. Papillomavirus has not been described in Atlantic offshore bottlenose dolphins; however, papillomavirus-like or papillomatosis infections have been reported in killer whales, harbor porpoise, Atlantic white-sided dolphins, narwhal, and beluga whales.

The offshore Atlantic bottlenose dolphin is a pelagic species that uses a different habitat and has a social structure different from that of coastal bottlenose dolphins. Exposure to most anthropogenic toxins/carcinogens is presumably less in pelagic species; however, certain airborne contaminants can have far-reaching effects and are known to persist in the environment. The cause of the neoplasms in this case is unknown. However, persistent pulmonary neomcdiosis may have induced neoplastic transformation in metaplastic epithelium because of chronic irritation, possibly in concert with a viral cohort or secondary to an inadequate immune response. Interestingly, pulmonary neomcdiosis was not observed at gross necropsy. Possible future investigation would include a search for a viral, toxic, or carcinogenic etiology, molecular genetic alteration, and immune status. This case represents the first report and characterization of pulmonary squamous cell carcinoma with metastasis in an offshore Atlantic bottlenose dolphin.

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Sources and manufacturers
a. Dako, Carpinteria, CA.
b. Becton Dickinson, Mountain View, CA.
c. Zymed, South San Francisco, CA.

References